SESQUITERPENE LACTONES IN A POPULATION OF BALSAMITA MAJOR CULTIVATED IN BULGARIA

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Abstract—Seven C-9 β -hydroxylated or esterified germacranolides were isolated from the flowers of a Balsamita major population cultivated in Bulgaria: the known 9 β -propionyloxy- and 9 β -isobutyryloxy-costunolides, 1 α ,10 β -epoxyhaageanolide, 9 β -hydroxyartemorin, and the new esters the propionyl-, isobutyryl- and 2-methylbutyryl-1 α ,10 β -epoxyhaageanolides. These lactones differ from the presently known eudesmanolides found in a B. major population cultivated in Poland, probably as result of the existence of chemotypes, frequently observed in the tribe Anthemideae.

INTRODUCTION

As part of our investigations on the sesquiterpene lactones in Bulgarian species of the Anthemideae [1, 2], we report here our findings on Balsamita major Desf†. This plant of Asian origin is widely grown in Europe and Asia. It is a large and up to 15 m high perennial plant with yellow flowers. Both flowers and leaves contain essential oils with carvone, thujone and β -cubebene as main components [4] Decoctions of leaves and flowers are used medicinally and against insects [5, 6]; this latter activity is due to the presence of pyrethrin I in the essential oil [4]. So far, only three selinane-type lactones, erivanin [7], isoerivanin and dehydroisoerevanin [8] have been reported for B major; they co-occur in a population from Poland.

RESULTS AND DISCUSSIONS

The material used was flowers of plants cultivated in the Botanical Garden of the Bulgarian Academy of Sciences, located near Sofia. The extraction procedure is described in the Experimental. Preparative chromatography on silica eventually yielded seven substances, six crystalline and one amorphous. All exhibited a strong IR band at 1750–1770 cm⁻¹ for a y-methylene lactone grouping proved further by ¹H NMR. Structure elucidation was carried out by means of ¹H NMR, EIMS and CIMS spectra compared with literature data.

All isolated compounds displayed several 250 MHz 1 H NMR signals with almost identical chemical shifts and multiplicity (Table 1), indicating their close structural relationships. Irradiation of the one-proton double doublet, located at δ 4.25–4.61 ($J = J_1 = 10$ Hz) and common for all compounds (1–7), assigned to H-6 by its coupling with one vinylic, H-5, and one trans axial, H-7, protons

Table 1. ¹H NMR spectra of sesquiterpene lactones 1–7 (250 MHz, CDCl₃, TMS as reference; chemical shifts in δ)

Н	1, 2	3–6	7
1	5 19 dd	2 85 dd	4 20 br d
$2\alpha,\beta$	2 32 m	1 46-2 29 m	1.97-2.26 m
$3\alpha,\beta$	2.32 m	1 46-2 29 m	1.97-2 26 m
5	4.69 d	5 20*, 5 24 br d	5 20 br d
6	4 57 dd	4 61 dd	4.25 dd
7	275 m	2 66*, 2.75† dd	276 m
8α)	2 05 m	2 25 br d	2.77 m
8B }		2 76 ddd	1.91 ddd
9	5 22 dd	3 21*, 4,39† dd	3 98 br d
13	6.31 d	6 30 d	6.22 d
13'	5 56 d	5.58*, 5 53 d	5 53 d
14	1 46 br s	1 15*, 1 25 s	5 38 br s
15	1 74 br s	1 85 s	1.59 br s
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MeCH₂COO (in 1 and 4): δ 1 17 (3H, dd, $J = J_1 = 7$), 2.40 (2H, m); (Me)₂CHCOO (in 2 and 5): δ 1 19 and 1 20 (2Me, two d, J = 6.9), 2.61 (H-2, m)

MeCH₂CH(Me)COO (in 6) δ 1 18 (3H, d, J = 7 1), 0.97 (3H, dd, J = J₁ = 7), 2 40 (H, m)

The δ values of 1, 2, 4-6 are on average within ± 0.05 ppm. The methylene envelope ($\delta 1.40$ -2.80) of all compounds was partially resolved by decoupling.

J(Hz) compounds 1 and 2, 1,2 α = 10.8, 1,2 β = 2.5, 5,6 β = 6 β ,7 α = 10, 7 α ,8 β = 9 5, 7 α ,13 = 3 5, 3 1 β ,2 β = 2 5; 1 β ,2 α = 5 6; 6 β ,7 α = 7 α ,8 β = 8 β ,9 α = 10, 7 α ,13 = 3 5, 4-6, 1 β ,2 β = 2, 1 β ,2 α = 11 1, 5,6 β = 6 β ,7 α = 7 α ,8 β = 10 1; 8 β ,9 α = 10.7, 7 α ,13 = 3 5, 7, 1 α ,2 β = 5 6, 6 β ,7 α = 7 α ,8 β = 8 β ,9 α = 10, 7 α ,13 = 3 5

The two one-proton doublets assigned for H-13 and H-13' (Table 1) confirmed the compounds as γ -methylene lactones. Irradiation of H-7 and H-9 demonstrated also the presence of two C-8 protons.

^{*}For compound 3

tfor compounds 4-6

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[†]Synonyms: Chrysanthemum balsamıta (L) Baillon, non L, Pyretrum majus (Desf) Tzvelev [3]

In addition to these common features, the ¹H NMR spectra of (1-7) contained signals characteristic for different substituents. The spectra of 1 and 2 displayed (a) a signal for one strongly deshielded carbinolic proton, FI-9, coupled with the two H-8, (b) two singlets for two olefinic methyls, (c) signal for one olefinic proton, H-1, coupled with one axial and one equatorial protons at C-2 The negative D_2O exchange experiments, the loss of 74 m/z (C_2H_5COOH) or of 88 m/z (C_3H_7COOH) from [M]⁺ of 1, $(m/z 304, C_{18}H_{24}O_4)$ or of 2 $(m/z 318, C_{19}H_{24}O_4)$ respectively, and the deshielded H-9 carbinolic proton, indicated 1 and 2 as C-9 esters. While 1 was clearly a C-9 propionate, the ¹H NMR signals for an isopropylic group indicated 2 as a C-9 isobutyrate. These data coincided well with those of the literature [8, 9] and identified (1) as the known 9β -propionyloxycostunolide, isolated from Inula royaleana [9], and 2 as the 9β -isobutyryloxycostunolide, found in Inula helenium [10]

The structures of 3-6 were elucidated in a similar way Instead of the characteristic ¹H NMR signal of the vinylic proton H-1 in 1 and 2 (Table 1), the compounds 3-6 showed a double doublet at $\delta 2.85 \pm 0.05$ for one proton coupled with a methylene group. The C-10 carbon bore a less deshielded tertiary methyl than that in 1 and 2 (δ 1 20 ± 0.05). The [M] $^+$ of (3) corresponded to $C_{15}H_{20}O_4$, but D₂O exchanged only one hydroxylic proton Hence, one oxygen in (3), and also in (4-6), formed a C-1 (10)-epoxide ring. In the ¹H NMR spectra of **4–6**, the carbinolic proton at the hydroxyl bearing C-9 displayed one deshielded double doublet at $\delta 439 \pm 0.05/\delta 321$ in 3 Their spectra also showed signals for protons in propionyl-, isobutyryland 2-methylbutyryl-acyls, forming the relevant C-9 esters. From these data, (3) is identified as the known 1α , 10β -epoxyhaageanolide, found in *Inula heterolepis* [9], while 4-6 appear to be the new C-9 esters of 3, 1e the 9β propionyloxy-, 9β -isobutyryloxy- and 9β -(2-methyl)butyryloxy-, 1α , 10β -epoxyhaageanolide respectively. The chemical shifts and the coupling constants of the protons in 4-6 coincided well with those of the parent compound (3) and showed that they shared a common stereochemistry

Instead of the C-10 methyl in compounds 1–6, compound 7, according to its ${}^{1}H$ NMR, contained one C-10 exomethylene group Two secondary hydroxyls in 7 were established by D₂O exchange and by the two broad doublets at δ 3 98 and δ 4.20 of two carbinolic protons The $[M+1]^{+}$ signal in CIMS, corresponding to $[C_{15}H_{20}O_4+H]^{+}$, split off two molecules of water and one of carbon monoxide This, and the ${}^{1}H$ NMR data in Table 1 identified 7 as the known 9β -hydroxyartemorin,

isolated earlier from *Inula heterolepis* and considered as a possible artefact of 3, both having been found together in the same *Inula* species [9]

As usual, the intensive signals with higher m/z in the CIMS (probe with isobutane) were more informative for the type of substituents in compounds 1-7 than the EIMS The CI spectra displayed also some peculiarities, for example as: (a) the esters with isobutyric, (2, 5), acids formed $[M]^+$ (100%) ions, (b) those with propionic (1 and 4), and with 2-methylbutyric (6) acids, as well as the alcohols (3 and 7) displayed $[M+1]^+$ [100%, 37% for (7)] ions, (c) certain differences in the intensity of the signals, produced by splitting of the acyl and/or water from $[M]^+$ or $[M+1]^+$ were also observed (Experimental) This could help in the identification and structure elucidation of similarly substituted lactones

As a contribution to the phytochemistry of Balsamita major Desf, one other finding of our work is that the Bulgarian population contained only C-9 β -hydroxylated germacranolides and their esters (7 compounds in total) In the chrysanthemum complex, only a few trans, trans-1(10),4(5)-germacradienolides have been found so far, and none with a C-9 hydroxyl [11] The lactones, known until now for Balsamita major Desf, are only of the eudesmane type [1, 2] and probably are specific for the Polish population Hence, this plant appears to occur in nature as more than one chemotype

EXPERIMENTAL

A voucher specimen of the plant has been deposited at the herbarium of the Institute of Botany, Bulgarian Academy of Sciences (c o Dr L Evstatieva), Sofia Mps uncorr IR in KBr pellets or in discs [comp (7)] ¹H NMR (Table 1) were recorded at WM 250 MHz in CDCl3 and TMS as ref MS by direct inlet, EI at 70 eV, CI with iso-butane at 200 eV. The air-dried and coarsely ground flowers (300 g) were extracted with 3 × 500 ml CHCl₃, evapd to dryness in vacuo, the residue stirred with 250 ml of 50% aq EtOH and defatted with petrol Precipitation of the aq EtOH part with Pb(OAc)2 in EtOH, filtration, removal of the EtOH from the filtrate in vacuo and re-extraction of the H2O residue with CHCl₃ yielded the crude lactone fraction (152 g) This material (40 g) was sepd on a prep-silica column by elution with CHCl3-Et2O mixtures The lactone fractions (IR monitoring) were further purified by prep TLC and recrystallization from Et₂O

9β-Propionyloxy costunolide (1) Colourless crystals, mp 143–145° (colourless oil acc [8]), $v_{\text{max}}^{\text{KBr}}$ cm⁻¹ 1760, 1720, 1660, 960, MS m/z, (rel int, %) EI 304 [M]⁺ (5), $C_{18}H_{24}O_4$, 248 (3), 230 [M – C_2H_3 COOH]⁺, (39), 57 [C_2H_3 CO]⁺ (100), CI – 305

 $[M+H]^+$ (100), 231 $[M+H-C_2H_5COOH]^+$ (70), 213, [231 $-H_2O]^+$ (8).

9 $\bar{\beta}$ -Isobutyryloxy costunolide (2). Colourless crystals, 152–154°; v_{max}^{KB} cm $^{-1}$ 1760, 1720, 1660, 1160, 1140, 960; MS· m/z, (%), EI 318 [M] $^+$ (6), C₁₉H₂₆O₄, 230 [M-C₃H₇COOH] $^+$ (30); CI· 318 [M] $^+$ (100), 231 [M-C₃H₇COO] $^+$ (60), 213 [231 -H₂O] $^+$ (3).

1α,10β-Epoxyhaageanolide (3). Colourless crystals, mp 136–139° (a gum acc. [9]), $v_{\text{max}}^{\text{KBr}}$ cm⁻¹ 3400, 1760, 1660, 1260, 1140, 950, 750, MS· m/z, (%), EI 246 [M – H₂O] + (1), 228 [M – 2H₂O] + (2), 149 (52), 81 (100); CI; 265 [M + H] + (100), 247 [M + H – H₂O] + (25), 229 [M + H – 2H₂O] + (10), 219 [247 – CO] + (3), 201 [229 – CO] + (1)

9 β -Propionyloxy-1 α ,10 β -epoxyhaageanolide (4). Colourless crystals, mp 180–183°; ν_{max}^{KBr} cm⁻¹ 1750, 1720, 1660, 1270, 1180, 1140, 970; MS. m/z (%), EI 320 [M]⁺ (1) $C_{18}H_{24}O_5$, 292 [M – CO]⁺ (0.5), 264 [M – 56]⁺ (1 5), 246 [M – C_2H_5 COOH]⁺ (2), 228 [246– H_2 O]⁺ (1 5), 57 [C_2H_5 CO]⁺ (100), CI. 321 [M + H]⁺ (100), 303 [M+H- H_2 O]⁺ (10), 247 [M+H- C_2H_5 COOH]⁺ (40), 229 [M+H- H_2 O- C_2H_5 COOH]⁺ (30)

9 β -Isobutyryloxy-1 α ,10 β -epoxyhaageanolide (5) Colourless crystals, mp 190–192° v_{mar}^{KBr} cm $^{-1}$. 1760, 1720, 1670, 1280, 1220, 1150, 960; MS, m/z, (%), E1 334 [M] $^+$ (2.5) $C_{19}H_{26}O_5$, 306 [M-CO] $^+$ (0.5), 264 (3), 246 [M $-C_3H_7COOH$] $^+$ (6); CI· 334 [M] $^+$ (100), 316 [M $-H_2O$] $^+$ (45), 247 [M $-C_3H_7COO$] $^+$ (50), 229 [M $-H_2O-C_3H_7COO$] $^+$ (38).

9 β -(2-Methyl)-butyryloxy-1,10-epoxyhaageanolide (6). Colourless crystals, mp 127–129°, $v_{\rm max}^{\rm KBr}$ cm $^{-1}$: 1760, 1720, 1660, 1280, 1210, 1145, 960, MS: m/z, (%), EI: 348 [M] $^+$ (2 5), $C_{20}H_{28}O_5$, 320 [M $^-$ CO] $^+$ (1), 291 [M $^-$ C $_4H_9$] $^+$ (1 5), 264 [M $^-$ 84] $^+$ (2), 246 [M $^-$ CQH] $^+$ (1 5), 85 (100), CI 349 [M $^+$ H] $^+$ (100), 331 [M $^+$ I $^-$ H $_2$ O] $^+$ (3), 247 [M $^+$ H $^-$ CQO] $^+$ (18), 229 [M $^+$ H $^-$ H $_2$ O $^-$ C $_4$ H $_9$ COO] $^+$ (10).

9β-Hydroxyartemorin (7). Oil as one TLC spot, partially crystallized after time. $v_{\rm max}^{\rm layer}$ cm⁻¹ 1760, 1670, 965, 262; MS: m/z, (%), EI. 246 [M-H₂O]⁺ (15) C₁₅H₁₈O₄, 228 [M-2H₂O]⁺ (18), 203 (22), 83 (90), 43 (100); CI: 265 [M+H]⁺ (37), 247 [M+1-H₂O]⁺ (15), 185 (8), 169 (15), 151 (27), 73 (100).

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REFERENCES

- 1. Ognyanov, I and Todorova, M (1983) Planta Med 48, 181
- 2 Todorova, M. and Ognyanov, I (1985) Planta Med 50, 174
- Heywood, V. H. (1976) in Flora Europea Vol IV, p 168 Cambridge Univ Press, Cambridge
- 4. Bestmann, H. J., Classen, B., Kobald, U., Vostrovsky, O. and Klingauf, F. (1986) Z. Naturforsch. 41C, 725.
- Davidov, B and Javashev, A. (1939) Materials for Bulgarian Botanical Dictionary, p. 134 Bulgarian Academy of Sciences.
- 6. Urumov, I. K (1935) Bulgarian Traditional Medicine, 2nd Edn. p. 11. Bulgarian Academy of Sciences. Sofia
- 7 Samek, Z., Holub, M, Bloszyk, E., Drozdz, B and Herout, V (1975) Coll. Czechosl. Chem Commun 40, 2676.
- 8 Samek, Z, Holub, M., Herout, V., Bloszyk, E and Drozdz, B (1979) Coll. Czech Chem Commun 44, 1468.
- 9. Bohlmann, F., Mahanta, P. K., Jakupovic, J., Rastogi, R. C. and Natu, A. A. (1978) Phytochemistry 17, 1165
- Bohlmann, F, Athes (Goren), N. and Grenz, M (1982) *Phytochemistry* 21, 1166
- 11 Seaman, P C (1982) Bot Rev 48, 123.